Towards better definition, quantification and treatment of fibrosis in heart failure. A scientific roadmap by the Committee of Translational Research of the Heart Failure Association (HFA) of the European Society of Cardiology


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Fibrosis is a pivotal player in heart failure development and progression. Measurements of (markers of) fibrosis in tissue and blood may help to diagnose and risk stratify patients with heart failure, and its treatment may be effective in preventing heart failure and its progression. A lack of pathophysiological insights and uniform definitions has hampered the research in fibrosis and heart failure. The Translational Research Committee of the Heart Failure Association discussed several aspects of fibrosis in their workshop. Early insidious perturbations such as subclinical hypertension or inflammation may trigger first fibrotic events, while more dramatic triggers such as myocardial infarction...
Introduction

Fibrosis is a fundamental process observed in cardiac remodelling and considered to be a key contributor to heart failure and its progression. Importantly, the presence and extent of myocardial fibrosis has also prognostic implications, as it causes contractile dysfunction and arrhythmias in structural heart disease of various aetiologies. Fibrosis is a direct and indirect target in the treatment of heart failure, either by established drug therapies (e.g. angiotensin-converting enzyme inhibitors or mineralocorticoid receptor antagonists) or specific anti-fibrotic drugs (e.g. pirfenidone). However, its resilience to therapy requires additional efforts to control (and ideally prevent or reverse) fibrotic remodelling, being identified as a major contributor to heart failure progression.

While fibrosis is a widely used term, the exact definition is less precisely defined. Fibrosis in the broadest sense is defined as excessive accumulation of extracellular matrix (ECM). In simplified terms, fibrosis can be divided into (i) ‘reparative fibrosis’ and (ii) ‘reactive fibrosis’. The development of an organized scar after myocardial infarction (MI) can be best described as reparative or replacement fibrosis, which is necessary to mechanically stabilize the evolving (necrotic) tissue defect. In contrast, the fine interstitial ‘reactive fibrosis’ encountered in non-ischaemic cardiomyopathies or in the surviving myocardium after MI appears to result from different pathological processes resulting in unique structural quality, ECM composition, and metabolic properties. Additionally, there is also a time component to scar development that has to be considered. For example, reactive fibrosis in the setting of pressure overload is initially characterized by perivascular fibrosis that later progresses to interstitial fibrosis. Fibrosis is also highly dynamic as it typically entails recruitment of fibroblasts and their conversion into myofibroblasts, excessive synthesis and secretion of ECM and ECM-associated modulatory glycoproteins, posttranslational modification and cross-linking of ECM proteins, and dysregulation of ECM production and breakdown by matrix metalloproteinases (MMPs) and their endogenous inhibitors (TIMPs). These different manifestations of fibrosis suggest that multiple targets or therapeutic opportunities may exist and that therapy may have to be personalized according to the diagnosis of specific remodelling processes and finally specific types of fibrosis.

Because today’s ‘one size fits all’ guideline approaches and broad heart failure patient classifications do not properly consider the different pathological processes that underlie fibrosis formation, it may not be a surprise that the outcomes of various studies are often contradictory and can only be rarely translated from one clinical setting to another. Also, as a consequence, fibrosis has not yet emerged as a primary target for heart failure therapies.

The Translational Research Committee of the Heart Failure Association (HFA) of the European Society of Cardiology (ESC) organized a workshop on myocardial fibrosis with the aim to discuss and recommend strategies to address knowledge gaps in this field. This scientific roadmap paper summarizes the principal knowledge gaps that were identified, including the need for (i) more specific definitions of processes underlying the formation of fibrosis in heart failure under different pathological conditions, (ii) improved methods to detect fibrosis using imaging techniques and biomarkers associated with specific entities of fibrosis; and (iii) new therapies to directly target specific processes underlying cardiac fibrosis. Here, we provide a framework to better define fibrosis during various stages, aetiologies, and severities of heart failure. We propose a structured experimental scheme to assess fibrosis quality as well as quantity, and to provide a work-up template that can be used in both translational and clinical research. Our goal is to direct future research to the identification of individual mechanisms of fibrosis formation, anticipating that this will provide insight into novel therapeutic targets and diagnostic tools for cardiac fibrosis stratification during heart failure progression.

More specific definitions are needed to describe the formation of myocardial fibrosis in heart failure

Myocardial fibrosis in heart failure is not a uniformly initiated process

Myocardial fibrosis is an endogenous, albeit suboptimal, repair response of the failing heart that can offer structural support while cardiomyocyte loss is occurring in the absence of appropriate cardiomyocyte replacement. Several key events characterize the fibrotic response to cardiac injury, which have been excellently reviewed elsewhere. Activation and conversion of fibroblasts into myofibroblasts are central events, critical also to non-cardiac fibrosis, being identified as a major contributor to heart failure progression. However, its resilience to therapy requires additional effor...
wound healing. Myofibroblasts produce and deposit ECM proteins, such as collagens, glycoproteins and proteoglycans (e.g. fibronectin, galectins, and periostin among many others), to offer local mechanical support to the failing heart. The formation of organized fibrotic structures and fibrils requires a multi-step process that involves the degradation and processing of existing ECM to remove damaged tissue, and the production, secretion, cross-linking, and maturation of new ECM. In addition to cardiac fibroblasts, which are the major source of ECM, monocytes and macrophages home to sites of injury and contribute to remodelling by secretion of pro-fibrotic growth factors. Finally, cardiomyocytes also contribute to secretion of pro-fibrotic growth factors into the ECM via paracrine mechanisms. In addition to fibrillar ECM constituents, non-structural glycoproteins and proteoglycans are important accessory mediators of fibrosis. Glycosylation is a highly prominent post-translational modification in ECM (reviewed by Rens et al.8) and glycoproteomics is a novel tool with promise in the study of myocardial fibrosis.8

Timely detection of fibrosis and determination of its state could potentially help to diagnose and stop heart failure progression early on. Capturing where fibrosis lies along the time continuum in a specific patient may inform physicians if fibrosis is developing as an early manifestation of the disease, and what type of targeted therapy could be employed. Early post-MI therapy will most certainly differ from the therapy of non-ischaemic diastolic dysfunction with a stiff left ventricle and even more from the treatment of end-stage heart failure with an often severely fibrotic myocardium. The exact differences in disease states, including the identification of disease modulators, must be identified to improve and ideally establish an individualized therapy of heart failure-related fibrosis.

Current classification of myocardial fibrosis

Traditionally, the form and stage of fibrosis have been denoted in line with the specific physiological phenomena that provoked the fibrotic response. Classically, the fibrotic process occurring after MI has been called ‘reparative’ or ‘replacement’ fibrosis. While scar formation is characterized by excessive accumulation of ECM, fibrosis is generally regarded as inevitable, as its absence would extend ventricular dilatation and could even result in ventricular rupture. Therefore, the infarct scar is a mandatory, albeit not perfect replacement, structural support. The post-MI scar is the result of dramatic cardiomyocyte loss and the subsequent deposition of collagen fibrils that are cross-linked to provide a strong ECM network. Reactive fibrosis, which is typically observed as perivascular or interstitial fibrosis, is stimulated by ongoing long-run maladaptive signalling (e.g. by inflammatory cells, paracrine signals, and oxidative stress) that is part of progressive pathological cardiac remodelling. Figure 7 summarizes the different types of fibrosis which may all be observed in parallel in the same heart, making it a challenging exercise to target distinct fibrotic processes. It depicts an example of histology from a virtual cardiac tissue biopsy — it becomes instantly apparent that even within the tissue same sample, different forms of fibrosis may be present, and thus, the chances for sampling error in real life are very real. Some fibrotic manifestations will be ‘reactive’, some will have been ‘reparative’, some will be (or have become) static, and others dynamic. It is quite often not feasible to distinguish the various forms, as many features are shared and transition into one another is possible. Thus, identification of the specific and probably dominant type of fibrosis will be important for individualized anti-fibrotic approaches.

Understanding myocardial fibrosis in different phenotypes of heart failure

Fibrosis is frequently described in experimental heart failure in preclinical animal models. Given its complex pathophysiology, it is crucial in such studies to provide a minimum amount of information to allow the reader understanding what fibrosis is referred to. Further, the triggers, dynamics, and characteristics of the fibrotic process are very different among various aetiologies of heart failure. Below, we discuss cardiac fibrosis in the setting of MI, pressure overload, and aging (Figure 2), as well as genetic cardiomyopathies and heart failure with preserved ejection fraction (HFP EF).

Myocardial fibrosis in post-myocardial infarction

Myocardial infarction is one of the most common causes of heart failure. Several animal models have been developed to model human MI. Permanent ligation of a coronary artery induces a large transmural MI (rat, mouse, dog, sheep, pig), while transient ligation causes ischaemia–reperfusion damage (mouse, rat, dog, pig), with variable degrees of damage depending on duration of ischaemia, selection of coronary artery, location of the ligature, and pre-treatment. MI causes a distinct tissue wound healing response with an initial strong inflammatory response, starting immediately after MI and peaking 3–7 days (depending on the species studied and model used) after MI. Neutrophils, monocytes, macrophages, but also fibroblasts themselves release factors that act on fibroblasts and trigger a pro-fibrotic response to form the infarct scar. The controlled invasion of inflammatory cells is a prerequisite for proper infarct healing and prevention of myocardial rupture.9–13 After the initial phase, inflammation subsides and the proliferative phase starts, where fibroblasts convert into myofibroblasts, migrate and proliferate, resulting in an increased capacity for wound contraction and repair. Additionally, fibroblast progenitors as well as endothelial to mesenchymal transition are considered important post-MI fibroblast sources. Collagen content begins to rise measurably 4–7 days after MI and peaks after 3–6 weeks, depending on the animal model used. Beside the amount of collagen, the type of collagen fibres formed and the degree of collagen cross-linking affect the mechanical properties of the tissue. Finally, a maturation phase is reached, where a stable scar is formed. During this phase, it is unclear whether the reduction in ECM turnover is due to reduced matrix synthesis, increased ECM breakdown, or both.

The precise role of fibroblasts and the fibroblast cell sources in the post-MI setting is incompletely understood, in part because...
of difficulties in labelling and identifying this cell type in vivo. Recently, fibroblast activating protein (FAP) was identified as a rather specific marker of activated, collagen-synthesizing fibroblasts, whereas inactive fibroblasts, or fully differentiated myofibroblasts and non-fibroblast cells in the infarct do not express FAP. Periostin is suggested as another marker of activated fibroblasts, suggestive for early fibroblast activation. Isolating post-MI cardiac fibroblasts from an in vivo-stimulated environment and evaluating these cells ex vivo has provided insight into their functional responses.

Myocardial fibrosis in models of pressure overload (e.g. hypertension)

The importance of fibrogenesis in pressure overload has been reviewed by Creemers and Pinto. Excessive myocardial ECM formation and collagen production take place in both human and experimental heart failure resulting from pressure overload, and collagen formation becomes disproportionate to left ventricular mass when the stress becomes chronic and sustained. Transforming growth factor (TGF)-β is a central protein in the formation of pressure overload-related fibrosis. It is activated by various circulating hormones such as angiotensin II and endothelin-1, but also by cellular stretch and strain. The TGF-β pathway leads to activation of Smad2/3 and Rho/ROCK signalling, and activation of stress-related kinases and proteins such as p38, ERK1/2 and elevated expression of connective tissue growth factor (CTGF). Fibroblasts in models of pressure overload have been identified as epicardial and endothelial cell-derived and Pax3-expressing cells (a major source under normal conditions and following pressure overload). Premature senescence of myofibroblasts was identified as an essential anti-fibrotic mechanism and potential therapeutic target in myocardial fibrosis in response to pressure overload.

Aging

Aging is one of the key drivers of myocardial fibrosis (reviewed in ). Animal models and human biopsy studies have demonstrated that collagen content of the heart progressively increase with advanced age, and collagen deposition is associated with increased wall stress, and with diastolic and systolic dysfunction. Various strategies have been developed to target the senescence factors in the heart, and other approaches have been evaluated in vivo, including genetic manipulation of the fibroblast population, pharmacological intervention, and even ablation techniques. These approaches may provide new insights into the mechanisms of fibroblast activation and senescence, and offer potential therapeutic strategies for the prevention and treatment of myocardial fibrosis.
systolic ventricular dysfunction. With aging, not only the production of collagen increases, but also the degradation becomes less effective.\textsuperscript{18,20,21} Also collagen processing and maturation is different, and cross-linking seems to increase.\textsuperscript{18,20,21} The triggers for fibrosis in the aging heart are manifold, and, as a result, fibrosis may present in multiple forms. In response to cardiomyocyte injury and cell loss, replacement fibrosis may be seen. At the same time, with ongoing inflammation and age-dependent increases in oxidative stress, interstitial fibrosis may occur. We must realize that age-dependent fibrosis will usually develop alongside, so in concert with fibrosis that develops in response to cardiac injury, which complicates the understanding of what causes and then supports sustained fibrotic processes.

**Myocardial fibrosis in (genetic) cardiomyopathies**

Fibrosis in (mono-) genetic cardiomyopathies can occur as fine interstitial fibrosis or replacement fibrosis, both due to structural changes in response to the gene defect. Therefore, the observation of fibrosis for instance on cardiac magnetic resonance imaging (MRI) is generally regarded as an early sign of the disease, even when systolic function is still normal.\textsuperscript{26,27} Early fibrosis in cardiomyopathies is regarded as a malicious event as the need for cardiac repair usually is minimal. Clearly, the events triggering fibrosis in cardiomyopathies are very heterogeneous, and encompass events such as cell death, metabolic derangements, neurohormonal activation, and direct toxic effects of mutated proteins.\textsuperscript{28}

**Myocardial fibrosis in heart failure with preserved ejection fraction**

Heart failure with preserved ejection fraction accounts for almost half of the cases of heart failure. Co-morbidities, including aging, obesity, hypertension, and diabetes, are key factors for HFpEF progression into overt heart failure. Recent evidence suggests that in HFpEF the extent of myocardial fibrosis (as measured by T\textsubscript{1}-MRI, see below) is related to the degree of diastolic dysfunction.\textsuperscript{29,30} Clearly, pro-fibrotic signals are diverse and differ from classical, systolic, heart failure signals. Fibrosis in HFpEF usually presents as perivascular and fine interstitial fibrosis and is associated with systemic inflammation.\textsuperscript{31} As a consequence, fibrosis in HFpEF will
likely be multifaceted, with fibrosis due to aging, due to hypertension, and in response to inflammatory and metabolic (obesity) triggers, with occasional superimposed reparative fibrosis, in case of (small) MI or myocarditis. Clearly, the current call to better phenotype HFrEF resonates particularly for the understanding of fibrosis in this complex disease.37

The ‘chronic fibrotic response’ in heart failure

From the discussion above it becomes clear that in heart failure (and in fact in more chronic diseases), a sustained fibrotic response is observed, that initially may be reparative, but at some point, rather contributes to organ damage and failure. So, it seems that in certain forms and stages of heart failure the fibrotic response cannot be switched off, and that a certain degree of fibrogenesis remains persistent. This is different from physiological healing, where the termination of the reparative phase is identified by the disappearance of activated myofibroblasts from the tissue.32,33

It is currently unknown how to differentiate the endogenous, necessary and beneficial fibrotic response or matrix turnover from the excessive, ongoing and harmful chronic fibrotic response that leads to matrix deposition and tissue stiffening.34 We postulate that these triggers that cause this chronic fibrotic response are multifold, including sustained fibroblast proliferation via feedback loops, cardiomyocyte-mediated fibroblast activation, inhibition of myofibroblast apoptosis, and the presence of sustained low-grade systemic and local inflammation.

Although mechanistically this remains largely a black box, several players have been recognized. As described above, TGF-β plays a central role in fibroblast proliferation and fibroblast-to-myofibroblast conversion. TGF-β is produced in high numbers by myofibroblasts to create a vicious cycle of myofibroblast activation. TGF-β stimulates several growth factors (epidermal growth factor, insulin-like growth factor-1, growth differentiation factor-11), which mediate proliferation of fibroblasts, involving autocrine signalling via fibroblast growth factor-2 and/or CTGF.35,36 TGF-β also prevents myofibroblast apoptosis, via stimulation of PI3K/AKT pro-survival signalling pathway.37 ‘Myofibroblast persistence’ may lead to non-resolving and progressive fibrosis, as exemplified by human idiopathic pulmonary fibrosis.38 Experimental drugs targeting the TGF-β and MAPK pathways indicate that the myofibroblast phenotype can be reversed, but whether this also can be achieved in vivo remains unclear.39,40

The low-grade, persistent systemic inflammation that is observed in heart failure is a major driver of fibrosis. TGF-β has pleiotropic effects on the immune system and has both immunosuppressive and pro-inflammatory functions,46 and may polarize macrophages and neutrophils towards a M2 phenotype, which produces large quantities of inflammatory cytokines.

Experimental studies suggest that regulating the inflammatory and immunomodulatory response may be effective in reducing MI-related remodelling, fibrosis and outcomes. For example, in a recent mouse study, neuregulin-1, an epidermal growth factor family member released by cardiac endothelial cells, attenuated myocardial interstitial fibrosis by inhibiting activation of myocardial macrophages.45 Most compelling evidence form the preclinical field was generated for tumour necrosis factor (TNF)-α inhibition43,46 but surprisingly, clinical studies e.g. with steroids and TNF-α blockers showed no or even detrimental effects, so that initial translation of experimental observations to clinical medicine has failed,44 suggesting that broad targeting of the immune system will not be a useful therapeutic strategy. Of interest, the recent CANTOS trial showed that targeted inhibition of interleukin-1α did reduce cardiovascular (and cancer) outcomes, albeit to a small degree.47

Proposal for a minimum assessment profile to screen cardiac fibrosis

The term ‘myocardial fibrosis’ needs to be specified with more precision and potentially individualized to offer effective therapeutics to patients with heart failure and cardiac fibrosis. For experimental studies, we propose a minimum set of parameters that should be provided to allow readers to appreciate the nuances of the fibrosis phenotype present. These items are listed in Table 1. In Table 2, we summarize the key functions, disease conditions, analytical methods and biomarkers in different forms of fibrosis. These insights are crucial to identify therapeutic opportunities for various subtypes of fibrosis-induced heart failure.

Improved methods are needed to detect fibrosis using biomarkers and imaging techniques

Detection of myocardial fibrosis is not straightforward in animal models and is even less so in the clinical setting where myocardial tissue sampling is not readily available. The gold standard for human studies is to assess the quality (i.e. focal, interstitial or perivascular distribution) and to quantify fibrosis in myocardial tissue biopsies using histological techniques (i.e. Masson’s Trichrome or Sirius Red histochemical staining). Although endomyocardial biopsies (EMBs) are limited by sampling error and small tissue fragments, the study of explanted hearts for heart transplantation offers unique possibilities with regard to spatiotemporal histological analyses and modern -omics techniques, not hampered by lack of tissue. We aware of several (national) initiatives, where all explanted hearts will be archived centrally to ensure proper sample size (so-called heart banks), and we foresee these will generate valuable information on fibrosis as well. But to date, tissue studies are mostly still carried out on EMBs, requiring an invasive procedure with the associated risk for complications and sampling errors.48 Thus, alternative, non-invasive and ideally equally or even more reliable methods should be developed and broadly applied.

Circulating biomarkers of myocardial fibrosis

Extracellular matrix proteins or cleaved processing products are often released into the systemic circulation and therefore measurable in serum or plasma using reliable and approved methods (e.g. ELISAs). Commonly used fibrosis biomarkers give insight into collagen production (e.g. procollagen type I N-terminal propeptide, procollagen type III N-terminal propeptide (PIIINP)) or the secretion of non-structural
Table 1 Proposed minimal dataset to describe fibrosis in animal studies

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Example(s)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Species</td>
<td>Mouse, rat, sheep, pig Precise reporting of strain, genetic background, age</td>
</tr>
<tr>
<td>Perturbation</td>
<td>Pressure overload, MI by permanent LAD ligation or ischaemia/reperfusion injury, diet, salt loading</td>
</tr>
<tr>
<td>Time course</td>
<td>Reporting the time course of disease progression, with samples taken before and at several time points [acute, subacute (days) and chronic (months)] post-disease induction</td>
</tr>
<tr>
<td>Assessments</td>
<td></td>
</tr>
<tr>
<td>Histology</td>
<td>For instance: Masson, Picrosirius red Percentage of LV tissue affected, sampled ROIs; Use of validated antibodies for immune histology</td>
</tr>
<tr>
<td>Inflammation glycoproteins-proteoglycans in the heart at RNA and protein level</td>
<td>Glycoproteins/proteoglycans: Periostin Osteopontin Syndecans Thrombospondins Osteoglycin TGF-β CTGF Galectin-3 Interleukin 1, -10, -11 Others pending on cardiac disease</td>
</tr>
<tr>
<td>Blood biomarkers</td>
<td>Galectin-3 CITP PIIINP ST2</td>
</tr>
<tr>
<td>Imaging</td>
<td>MRI (T1 mapping, late enhancement fibrosis)</td>
</tr>
<tr>
<td>Functional analyses</td>
<td>Echocardiography and invasive haemodynamics for determining load-dependent diastolic and systolic function</td>
</tr>
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</table>

CITP, C-terminal propeptide of procollagen type I; CTGF, connective tissue growth factor; LAD, left anterior descending artery; LOX, lysyl oxidase; LV, left ventricular; MI, myocardial infarction; MMP, matrix metalloproteinase; MRI, magnetic resonance imaging; PIIINP, procollagen type III N-terminal propeptide; ROI, region of interest; TGF, transforming growth factor; TIMP, tissue inhibitor of metalloproteinases.

(glyco)proteins that modulate the collagen production itself or its maturation (e.g. periostin, mimecan, monocyte chemoattractant protein-1, or galectin-3). In addition, several MMPs (e.g. MMP-3, MMP-9, MMP-11, and MMP-12) and their tissue inhibitors (e.g. TIMP-1 and TIMP-3) involved in the balance of collagen degradation, are released into the blood stream and can be measured reliably. In fact, there is a wide body of literature on the potential utility of these markers alone or in concert.49 In general, many of these markers are valuable for clinical risk prediction. Interestingly, several factors have been shown to predict the response to treatment with anti-fibrotic properties, such as mineralocorticoid receptor antagonists.50 However, the use of these markers has not become a clinical standard because of limited power in fully adjusted models with clinical variables, and because of technical difficulties in measuring the proteins, often requiring laborious and expensive radioimmunoassays. Recently, several new emerging fibrotic markers have been studied, including galectin-3, sST2, and periostin.51 These markers can generally be measured with Food and Drug Administration-cleared ELISA assays which allow fast turnaround times.

Circulating levels of fibrosis markers may not parallel findings in histologically proven cardiac fibrosis and therefore caution is required in the interpretation of systemic venous circulating biomarker levels in relation to myocardial disease. López et al.52 measured circulating levels of many fibrotic markers and correlated these in the same patients to local cardiac tissue fibrosis volumes measured with histology. The results show that out of 28 potential biomarkers associated with...
<table>
<thead>
<tr>
<th>Type of fibrosis</th>
<th>Function</th>
<th>Disease conditions</th>
<th>Methods to detect in tissue</th>
<th>Blood biomarkers</th>
</tr>
</thead>
<tbody>
<tr>
<td>Reparative fibrosis</td>
<td>Replacing necrotic cells</td>
<td>Myocyte necrosis (ischaemia, infection, autoimmunity, toxicity, gene mutations)</td>
<td>Histochemistry (Sirius red and Azan blue)</td>
<td>CITP, PICP, PINP, PIINP</td>
</tr>
<tr>
<td>Reactive fibrosis</td>
<td>New matrix production in between cells</td>
<td>Matrix production in response to an acquired or genetic trigger, such as stretch</td>
<td>Cardiac MRI, late enhancement</td>
<td>CITP, PICP, Galectin-3, sST2, and peristin</td>
</tr>
<tr>
<td>Perivascular fibrosis</td>
<td>New matrix produced around vessels</td>
<td>Perivascular fibrosis upon pressure overload or vascular stress</td>
<td>Histochemistry</td>
<td>Unknown</td>
</tr>
</tbody>
</table>

Cells involved
- (Myo)fibroblasts
  - Reparative and reactive
    - Most diseased hearts
      - Staining of peristin, FAP, SMA
      - Peristin, mimecan, SPARC
- Smooth muscle cells
  - Perivascular and reactive
    - Pressure overload, vasculitis
      - Staining of SMA
- Inflammatory cells
  - Reparative and reactive
    - Acute and chronic cardiac disease
      - Staining of macrophages, monocytes, neutrophils (CD45)

Content/mechanisms
- Collagen production
  - Structural proteins
    - Healthy and diseased heart
      - Immunostaining, rtPCR
      - CITP, PICP, PINP, PIINP
- Glycoproteins/proteoglycans
  - Inter-cellular communication
    - Most diseased hearts
      - Immunostaining, rtPCR
      - Galectin-3, sST2, peristin, mimecan, SPARC
- MMP and their inhibitors (TIMPs)
  - Collagen degradation/production balance
    - Healthy and diseased heart
      - Immunostaining and activity assays, rtPCR
      - MMP-1 and -9, TIMP-1 and -2
- Growth factors: TGF, CTGF, FGF, Wnt pathways
  - Stimulate collagen production, affects inflammation
    - Most diseased heart
      - Immunoblots, signalling pathways
      - ND

CITP, C-terminal propeptide of procollagen type I; CRP, C-reactive protein; MMP, matrix metalloproteinase; MRI, magnetic resonance imaging; PINP, amino-terminal propeptide of type I collagen; PIINP, procollagen type III N-terminal propeptide; PIIINP, procollagen type I N-terminal propeptide; rtPCR, reverse transcriptase-polymerase chain reaction; sST2, soluble ST2; TIMP, tissue inhibitor of metalloproteinases.
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Figure 3 Systemic biomarkers, measured in the plasma of patients with heart failure, ideally reflect changes in the heart muscle. For cardio-specific biomarkers, such as natriuretic peptides and troponins, this is very accurate. However, for many (more novel) markers that are expressed by many organs outside the heart as well, the systemic levels only marginally reflects cardiac production. BNP, B-type natriuretic peptide. (Illustration: Maartje Kunen, Medical Visuals.)

build-up or breakdown of myocardial fibrosis, only C-terminal propeptide of procollagen type I (CITP) and PIIINP correlated with histological findings. This may be explained because in heart failure patients, several other organs undergo fibrotic changes as well: liver, lungs, kidneys, and vessels. A recent article by Du and colleagues showed that galectin-3, growth differentiation factor-15, and TIMP-1 plasma levels do not reflect myocardial fibrosis in mouse models of post-MI heart failure, hypertensive heart failure, and HFpEF. Instead, production in extracardiac tissues such as fatty and lung tissue had much greater impact on plasma levels of these markers. So ideally, we would need a marker that is specific for cardiac fibrosis, or at the very least, adequately reflects changes in myocardial fibrosis. However, since most fibrotic pathways are shared amongst organs, such a marker may not exist and it is therefore simply impossible to use circulating fibrosis markers as a perfect surrogate for myocardial fibrosis. Further, the markers generally do not clearly distinguish between various forms of fibrosis, or between the trigger that causes fibrosis. Therefore, the use of these factors for precision diagnostics is questionable, but excess local production however may be used to target specific treatment. Further, the potential of fibrotic markers as surrogate outcomes in phase I/II trials is likely to be limited at this point, in view of the limited specificity of available fibrotic (bio-)markers.

Imaging techniques to visualize and quantify fibrosis

The most widely applied imaging technique in contemporary heart failure management is echocardiography. Classical two-dimensional echocardiography, however, provides little information about the presence or extent of fibrosis. More modern techniques such as tissue velocity imaging and global longitudinal systolic strain provide mechanical tissue details that associate with myocardial fibrosis by biopsy examination. In addition to echocardiography, cardiac MRI is a technique that is replacing EMB as gold standard for human cardiac fibrosis identification and quantification. Clearly, the use of cardiac MRI cannot fully replace echocardiography as first-choice imaging modality, given the high costs. But in order to perform in deep phenotyping, with the aim to choose patients with specific pathophysiological characteristics, or to monitor these during (drug) treatment, cardiac MRI has distinct benefits. Using delayed gadolinium enhancement, it is possible to visualize scar tissue, for instance after transmural myocardial infarction. While delayed gadolinium enhancement mainly identifies focal reparative fibrosis, modern techniques in cardiac MRI are developing that may be able to provide more granularity in imaging fibrosis. Most interestingly, T1 mapping is such an emerging cardiac MRI technique, measuring the longitudinal relaxation time of individual protons, which is depicted as a pixelated map. T1 mapping allows the quantification of extracellular volume (ECV) fraction of the myocardium. ECV is not a pure measure of fibrosis, although it has been evaluated to this aim, but ECV rather mirrors diffuse changes including fibrosis, but also interstitial oedema, protein degradation and aggregation, lipid accumulation, and deposition of iron or amyloid. A recent HFA position paper discusses in detail the (fast) developments in imaging techniques. Furthermore, the presence and the extent of MRI-proven fibrosis have been related to poor clinical outcomes. Collectively, different imaging (MRI) techniques may be applied as surrogate measures for the presence and extent of myocardial fibrosis. The next challenge is to develop a therapeutic plan to reverse or prevent further development of fibrosis based on the cardiac magnetic resonance findings. A promising approach would be to label specific molecules with established relation to myocardial fibrosis, and image
that are on the horizon.

appointing result thus far. Next, we will discuss a few novel options important in MMP inhibition and this could in part explain the disappointing results thus far. Next, we will discuss a few novel options that are on the horizon.

New therapies are needed to directly target myocardial fibrosis with specific drugs

It has been proposed for several years that direct targeting fibrosis might be useful in heart failure. Angiotensin-converting enzyme inhibitors and mineralocorticoid receptor antagonists reduce fibrosis formation, but clearly a residual fibrotic burden remains and with that, a potential need to target it in order to improve outcomes. It is crucial to ascertain where the fibrotic process is, at a given time point, what the triggers are, and which cells and proteoglycans play a role. The complexity of targeting fibrosis is illustrated by a recent article by Clarke and colleagues, providing an example for why MMP inhibition may not be as effective as previously hypothesized. They postulate that early during the remodelling phase, MMP inhibition might be less effective because there is little collagen to degrade, while at later fibrotic phases it is less effective because MMP levels have fallen to low levels. Therefore, the setting, the aetiology and the timing, all appear very important in MMP inhibition and this could in part explain the disappointing results thus far. Next, we will discuss a few novel options that are on the horizon.

Connective tissue growth factor and pirfenidone

Inhibition of fibrosis formation in pressure overloaded heart is best achieved by alleviating the primary stressor, i.e. the elevated pressure. TGF-β (e.g. pirfenidone), angiotensin or endothelin receptor blockers, ERK inhibitors or inhibition of CTGF are being tested as specifically fibrosis targeted treatments. Different from post-MI fibrosis, it appears that inhibition of the excess fibrosis in pressure overload is generally safe and well-tolerated.

Connective tissue growth factor (or CCN2) is a matricellular protein and modulates the signalling of many cytokines and ECM signals including those of TGF-β, bone morphogenetic protein, Wnt, vascular endothelial growth factor, and integrins. Because it modulates multiple pathways simultaneously, and via several different mechanisms, the effects of CTGF are combinatorial and context-dependent (i.e. dependent on the environment and mediators present), and therefore, the biology of CTGF is very complex. CTGF expression is induced by many different pathophysiological insults, and when it becomes overexpressed, it helps promote differentiation of cells to become activated myofibroblasts that deposit and remodel the ECM. CTGF is involved in multiple positive feedback loops that can propagate tissue remodelling and fibrosis, and therefore it should be considered a central mediator of fibrosis. Consequently, the goal of inhibiting CTGF is to disrupt these positive feedback loops and arrest the progressive nature of fibrosis (and possibly reverse it). A human monoclonal antibody, FG-3019, that binds to CTGF and interferes with its activity has proven beneficial in rodent models of MI (unpublished), neonatal (rat) bronchopulmonary dysplasia, and thoracic aorta constriction model. Clinical testing of FG-3019 indicates an excellent safety profile and has been tested in approximately 400 patients with diabetic kidney disease, pancreatic cancer, idiopathic pulmonary fibrosis, or liver fibrosis.

Pirfenidone is a synthetic molecule that has been reported to decrease the expression of various pro-fibrotic factors, including TGF-β1, TNF-α, platelet derived growth factor and collagen. Results from experimental models provided evidence for a therapeutic utility of pirfenidone in pressure overload and hypertension, leading to a reduction in arrhythmogenic substrate.

Matrix metalloproteinases

The effects of regulating MMP activity on cardiac fibrosis and left ventricular remodelling outcomes have primarily been assessed in MI models. Targeted deletion and transgenic mice or MMP inhibitors (MMPI) reveal both beneficial and detrimental consequences. While early promises of MMPI in animal models were encouraging, these findings have not translated to humans. This has been due to selectivity and specificity issues, as well as our lack of understanding of the full range of MMP functions. For example, the MMP-9 substrate list includes hundreds of substrates ranging from collagen and fibronectin, interleukin-1β, pro-enzymes and citrate synthase. Further, not all MMPI have been beneficial, as MMPI-12i given at 3 h post-MI suppressed neutrophil apoptosis to prolong inflammation, resulting in exacerbated left ventricular dilatation. MMP-28 deletion inhibited M2 anti-inflammatory macrophage activation to stimulate left ventricular dysfunction and increase cardiac rupture rates. If anything, these results indicate how complex the fibrotic process in heart failure is, and the specific role of MMPs herein. There is a need, therefore, to delineate individual MMP roles under specific conditions and times.

Galectin-3 inhibitors

Galectin-3 is a lectin binding galactoside, and has been shown to be upregulated in heart failure by myofibroblasts, monocytes and macrophages that are recruited towards sites of injury and fibrosis. Studies in mice deficient for galectin-3 have suggested that galectin-3 is not a bystander but rather a culprit for myocardial fibrosis. Inhibition of galectin-3, either achieved with large carbohydrates, or antisense RNA or small designer molecules, effectively reduces organ fibrosis.

Non-coding RNAs

Recently, non-coding RNA (both microRNA and long non-coding RNAs-based) based treatment strategies for fibrosis have been put forward. Specific non-coding RNAs seem to play crucial roles in the regulation of the cardiac fibroblast phenotype and their modulation seem to be effective both in animal as well as clinical studies; indeed there is currently a phase II trial in patients with kidney fibrosis using an inhibitor of microRNA-21
Future directions and conclusions

In this article we discuss several challenges and requirements in the study of fibrosis (Table 3). For future applications, strategies that allow for differentiation of fibrosis subtype are needed. Preclinical studies in relevant animal models to better understand the dynamics of fibrosis formation, degradation, and their importance for the functional phenotype are therefore required. More advanced in vitro models might allow a better functional control and mechanistic understanding. Whereas further optimizations are needed to implement human cardiac disease parameters, these well controlled environments allow longitudinal evaluations and screening of anti-fibrotic strategies. Accumulating data linking fibrosis to a stronger inflammatory response are at the initial phases. At later time points, fibrosis-specific mediators and pathways (predominantly fibroblast-specific factors such as TGF-β, osteopontin, and galectins) contribute to the progression of fibrosis, and are distinct from the mechanisms driving inflammation. The presence of co-morbidities should be considered — e.g. it has recently been discussed that co-morbidities such as cancer may obscure the biomarkers’ signals. Thus, to design effective therapeutics for fibrotic disease, inflammation triggering fibrosis is to be considered, and the challenge ahead is to target specific molecules and pathways that act on fibrosis (specific interleukins) while leaving the (often beneficial) effects of the inflammatory response uninhibited. A vast diversity of inflammatory, immunological, and molecular mechanisms collectively contribute to cardiac fibrosis: the complex interplay between adaptive immune system activation, fibroblasts-to-myofibroblast conversion and proliferation, mast cell activation, neutrophil influx, and production, modulation, maturation and apposition of collagens, the embedding in the extracellular milieu. These should all be considered and taken into account during the design and testing of new anti-fibrotic therapies.

Clinically, disease-specific (bio-)markers and imaging modalities — acting as surrogate parameters of specific temporal stages of fibrosis — will help to identify patients who might benefit from a specific therapy. Until recently, attempts to inhibit fibrosis have been mostly focusing on single pro-fibrotic factors. Since fibrosis is driven and sustained by the activation of multiple interconnecting and intercommunicating pro-fibrotic pathways, a multi-target approach will likely help to slow down the progression of fibrosis. Using systems biology approaches and multi-omics technologies to understand network signalling will aid in these efforts. Ultimately, a concerted anti-fibrotic strategy that collectively targets important inflammatory signalling molecules, pro-fibrotic cytokines, and cellular functions should be considered in developing therapies to adequately treat fibrosis.

In conclusion, in this position paper we have summarized the current knowledge gaps in the myocardial fibrosis field and provided templates for assessing fibrosis in both translational and clinical studies.

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References


